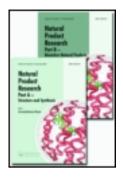
This article was downloaded by: [University of Nevada - Reno] On: 13 November 2012, At: 14:51 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Natural Product Research: Formerly Natural Product Letters

Publication details, including instructions for authors and subscription information: http://www.tandfonline.com/loi/gnpl20

In vitro activity of compounds isolated from Piper crassinervium against Trypanosoma cruzi

Adriana Aparecida Lopes^a, Silvia Noelí López^a, Luis Octávio Regasini^a, João Marcos Batista Junior^a, Daniela Luz Ambrósio^b, Massuo Jorge Kato^c, Vanderlan da Silva Bolzani^a, Regina Maria Barretto Cicarelli^b & Maysa Furlan^a

^a Instituto de Química, Universidade Estadual Paulista, C.P. 355, CEP 14801-970, Araraquara, SP, Brazil

^b Faculdade de Ciências Farmacêuticas, Universidade Estadual Paulista, CEP 14801-902, Araraquara, SP, Brazil

^c Instituto de Química, Universidade de São Paulo, C.P. 26077, CEP 05599-970, São Paulo, SP, Brazil

Version of record first published: 10 Sep 2008.

To cite this article: Adriana Aparecida Lopes, Silvia Noelí López, Luis Octávio Regasini, João Marcos Batista Junior, Daniela Luz Ambrósio, Massuo Jorge Kato, Vanderlan da Silva Bolzani, Regina Maria Barretto Cicarelli & Maysa Furlan (2008): In vitro activity of compounds isolated from Piper crassinervium against Trypanosoma cruzi, Natural Product Research: Formerly Natural Product Letters, 22:12, 1040-1046

To link to this article: <u>http://dx.doi.org/10.1080/14786410802243271</u>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <u>http://www.tandfonline.com/page/terms-and-</u> conditions

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary

sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



In vitro activity of compounds isolated from Piper crassinervium against Trypanosoma cruzi

Adriana Aparecida Lopes^a, Silvia Noelí López^a, Luis Octávio Regasini^a, João Marcos Batista Junior^a, Daniela Luz Ambrósio^b, Massuo Jorge Kato^c, Vanderlan da Silva Bolzani^a, Regina Maria Barretto Cicarelli^b and Maysa Furlan^{a*}

^aInstituto de Química, Universidade Estadual Paulista, C.P. 355, CEP 14801–970, Araraquara, SP, Brazil; ^bFaculdade de Ciências Farmacêuticas, Universidade Estadual Paulista, CEP 14801–902, Araraquara, SP, Brazil; ^cInstituto de Química, Universidade de São Paulo, C.P. 26077, CEP 05599–970, São Paulo, SP, Brazil

(Received 15 December 2007; final version received 18 June 2008)

This study describes the antichagasic potential of five compounds isolated from leaves of *Piper crassinervium* (Piperaceae). Two prenylated benzoic acid derivatives, one prenylated hydroquinone and two flavanones, were evaluated. The *in vitro* trypanocidal activity was determined against epimastigote forms of *Trypanosoma cruzi* (Y strain), the etiologic agent of Chagas disease. The most active compound was the prenylated hydroquinone [1,4-dihydroxy-2-(3^0 ,7⁰-dimethyl-1⁰-oxo-2⁰-*E*,6⁰-octadienyl)benzene] with an IC₅₀ value of 6.10 µg mL⁻¹, which was in the same order of activity if compared with the positive control benznidazole (IC₅₀ = 1.60 µg mL⁻¹). This is the first report of trypanocidal activity for prenylated hydroquinone and benzoic acid derivatives.

Keywords: prenylated benzoic acid; prenylated hydroquinone; flavanone; *Trypanosoma cruzi*; *Piper crassinervium*

1. Introduction

American trypanosomiasis or Chagas disease is a zoonosis caused by the hemoflagellate protozoan *Trypanosoma cruzi*. In Latin and South America millions of people (estimated at 16–18 million in the decade of the 1990s) have been infected, and more than 25% of the population is at risk of being contaminated by the parasite (Grael, Albuquerque, & Lopes, 2005). However, over the last 15 years, prevalence estimates have been reduced, largely due to intensive large-scale programs designed to halt transmission by eliminating domestic populations of the insect vectors and to improve the screening of blood donors (Jannin & Villa, 2007). Nevertheless, transmission of Chagas disease is still a major health problem because of the transfusion of infected blood. Moreover, a recent case report in Brazil suggested the oral transmission of parasites, renewing the discussions about this problem (Pérez-Gutiérrez, Agrelo Salvatella, & Figueroa, 2006). Currently available chemotherapy, based on nifurtimox and benznidazole, is accepted for treating acute infections, but it is unsatisfactory in the prevalent chronic stage because of limited efficacy, besides the

^{*}Corresponding author. Email: maysaf@iq.unesp.br

frequent toxic effects (Luize, Ueda-Nakamura, Dias Filho, Garcia Cortez, & Nakamura. 2006). At present, the only trypanocidal compound used to prevent infection in blood banks is gentian violet, but it shows toxic effects and its use is limited (Rosas et al., 2007). The lack of effective medicines against acute and chronic infection is the critical reason to find more efficient and less toxic drugs than those currently used on the way to a cure for this pan-Latin-American disease. In spite of the great advances in modern medicine in recent decades, plants still make an important contribution to health care, providing chemical diversity and bioactivity, as well as to the development of hundreds of pharmaceutical drugs (Luize et al., 2006). Piperaceae family, belonging to the largest basal Angiosperm order, includes two rich genera, Piper (ca. 2000 species) and Peperomia (ca. 1500–1700 species) (Wanke et al., 2007). Several Piper species have been well studied, in which *Piper nigrum* produce piperine, the active pungent principle of black pepper; from P. hispidum, P. arboreum and P. tuberculatum were isolated antifungal and insecticide amides (Navickiene et al., 2000; Silva et al., 2002). Additionally, chromenes and polyketides with significant antifungal activity have also been isolated from P. aduncum, P. hostmanianum, P. crassinervium and P. gaudichaudianum (Baldoqui et al., 1999; Danelutte, Lago, Young, & Kato, 2003; Kato & Furlan, 2007).

Hydroquinones bearing prenyl moieties are natural products uncommon to land plants, but highly frequent in Piper species (Lago et al., 2004). Reported biological activities for such compounds include antitumoral (Danelutte et al., 2003), analgesic (De Pasquale, Circosta, Occhiuto, De Rosa, & De Stefano, 1991), antioxidant (Yamaguchi, Lago, Tanizaki, Di Mascio, & Kato, 2006) and antimicrobial (Lago et al., 2004). Prenylated benzoic acid derivatives were also isolated from *Piper* species, including P. crassinervium, and their bioactivity spectra are similar to prenylated hydroquinones. Here we report the evaluation of trypanocidal activity for five known compounds, 1 [4-hydroxy-(3',7'-dimethyl-1'-oxo-octa-2'-E-6'-dienyl)benzoic acid] (Lago et al., 2004), **2** [4-hydroxy-3-(3',7'-dimethyl-3'-hydroxy-1'-oxo-6'-octenyl)benzoic acid] (Lago et al., 2004), **3** [1,4-dihydroxy-2-(3',7'-dimethyl-1'-oxo-2'-E-6'-octadienyl)benzene] (Danelutte et al., 2003), 4 isosakuranetin (5,7-dihydroxy-4'-methoxyflavanone) (Agrawal & Bansal, 1989) and 5 sakuranetin (5,4'-dihydroxy-7-methoxyflavanone) (Agrawal & Bansal, 1989; Danelutte et al., 2003), (Table 1) isolated from leaves of *P. crassinervium*, as a part of our program devoted to discover new hits in the development of drugs for treatment of Chagas disease. This constitutes the first report of trypanocidal activity for prenylated hydroquinone and benzoic acid derivatives.

2. Materials and methods

2.1. Plant material

Leaves of *P. crassinervium* Kunth were collected in the region of Vale do Ribeira, Atlantic Forest (São Paulo State, Brazil) and identified by Dr. Guillermo E.D. Paredes (Universidad Nacional Pedro Ruiz Gallo, Peru). A voucher specimen (KATO-0084) was deposited at the Herbarium of Instituto de Botânica, São Paulo, SP, Brazil.

2.2. Extraction and isolation of the constituents

Dried leaves (350 g) were extracted with EtOAc and concentrated to dryness under vacuum. The crude extract (20 g) was resuspended in MeOH: H₂O (8:2) and partitioned

Compounds	Chemical structure	$IC_{50} \ (\mu g m L^{-1})$
1	OH O	16.3
2		15.8
3	OH OH	6.1
4		119.6
5	O O O OH	111.7
Benznidazole ^a		1.6

Table 1. Inhibition of epimastigote forms of *T. cruzi* by compounds 1–5 isolated from *P. crassinervium.* IC_{50} values are given in μ g mL⁻¹.

Note: ^aPositive control.

with hexane and EtOAc. The hexane soluble portion (6.0 g) was fractioned using column chromatography over silica gel (70–230 mesh; Merck, column size 3.0×25 cm), which was eluted with hexane containing increasing amounts of EtOAc (up to 30%) to yield 11 fractions. Fraction 4 (0.284 g) was further separated by column chromatography over reversed phase ODS (Merck; C-18; 1.0×4.5 cm column) with MeOH–H₂O (4:1) to yield eight fractions; the first fraction (0.221 g) was submitted to column chromatography over silica gel (Merck; 230–400 mesh; 1.0×3.0 cm column) eluted with hexane containing increasing amounts of EtOAc (up to 30%) to yield six fractions, the third fraction yielded compound **3** (0.023 g). Fraction 6 (0.015 g) was applied to a column chromatography over reversed phase ODS (Merck; C-18; 1.0×4.5 cm column) with MeOH–H₂O (4:1) to yield compound **1** (0.011 g).

The portion soluble in EtOAc (3.0 g) was fractioned by column chromatography over LH-20 (Merck, column size 3.0×100 cm) and eluted with MeOH to yield 16 fractions. Fraction 3 (0.050 g) was submitted to preparative TLC over silica gel (230–400 mesh; Merck), which was developed with hexane–acetone (6:4) to yield compound 2 (0.009 g). Fraction 11 (0.100 g) was chromatographed on preparative HPLC using hexyl-phenyl phase, which was eluted with MeOH–H₂O (7:2), to yield 4 (0.030 g) and 5 (0.010 g). Compounds 1–5 were identified by comparison of their physical properties with literature values.

2.3. Trypanocidal assay

The assays of the anti-parasitic activities of the isolates were conducted by screening their effects on the proliferation of epimastigote forms of *T. cruzi* Y-strain, grown in axenic culture in LIT medium. Stock solutions (containing 1.0 mg mL^{-1}) of all tested compounds were prepared in dimethylsulphoxide and serially diluted (1:3) in liver infusion tryptose medium to yield sample solutions with concentrations of 0.41, 1.23, 3.70, 11.1, 33.3, 100.0 and 300.0 µg mL⁻¹. All assays were conducted in triplicate. The number of remaining viable protozoa was established by counting in a Neubauer chamber following incubation at 28°C for 72 h with the test sample. The 50% inhibitory concentration (IC₅₀) values and statistical data were determined by using Probit analysis statistical method with GW-BASIC 2.33 (Microsoft Software, 1987). The IC₅₀ values for compounds 1–5 and for benznidazole (employed as positive control) are shown in Table 1.

3. Results and discussion

In this work, we have tested the inhibitory effects of five known compounds isolated from *P. crassinervium* against epimastigote forms of *T. cruzi*. Results are presented in Table 1. Three groups of molecules were tested here, two benzoic acid derivatives (1 and 2), one hydroquinone derivative (3), and two flavanones (4 and 5). Moreover, compounds 1-3 are *C*-geranylated molecules.

The results showed that geranylated compounds were more active than the flavanones. Within the active compounds, the best activity was observed for 3 (IC₅₀ value of 6.10 μ g mL⁻¹) followed by geranylated *p*-hydroxybenzoic acid derivatives 1 and 2. The IC₅₀ value observed for compound 3 [1,4-dihydroxy-2-(3',7'-dimethyl-1'-*oxo*-2'-*E*-6'-octadienyl)benzene], a *p*-hydroquinone derivative, represents an interesting hit

if compared with positive control benznidazole (IC₅₀ value $1.6 \,\mu g \,m L^{-1}$, Table 1). This molecule has two phenolic hydroxyl groups, one of them with a chelated function, which is also present at compounds 2 and 3. The change of one phenolic hydroxyl group (compound 3) by a carboxylic acid one (compound 1) produced a loss of activity greater than 50% (IC₅₀ value from 6.1 to $16.3 \,\mu g \,m L^{-1}$, Table 1), suggesting its crucial role in trypanocidal activity. This observation is in agreement with Luize, P.S. et al., who proposed that the loss of the hydroxyl group in neolignans isolated from *P. regnellii* resulted in the complete loss of activity against epimastigote forms of *T. cruzi* (Luize et al., 2006). In addition, the phenolic hydroxyl group present in the active molecule is apparently able, through hydrogen-bond links with the specific molecules on the cell surface, to inactivate essential metabolic pathways within the parasite (Patrick, 1995). The decrease in the activity observed for compound 2 (IC₅₀ value 15.8 $\mu g \,m L^{-1}$, Table 1), is also in agreement.

Isosakuranetin (4) and sakuranetin (5) (Agrawal & Bansal, 1989; Danelutte et al., 2003), showed weak antichagasic activity with IC_{50} values of 119.6 and 111.7 µg mL⁻¹, respectively (Table 1). Several types of flavonoids have been identified as antiprotozoal principles of plant extracts (Camacho et al., 2002; Grael et al., 2005; Jordão, Vichnewski, Souza, Albuquerque, & Lopes, 2004). However, there are few reports about the activity of flavonoids against epimastigotes of *T. cruzi*. Our results are in agreement with such observations, suggesting that the molecular topology is very important for adequate interactions at receptor level to exert trypanocidal effect. Flavanone skeleton was less active than geranylated hydroquinone derivatives, even with phenolic hydroxyl groups in the molecule.

The presence of lipophilic geranyl moiety oxygenated in benzylic position, in compounds 1-3 could be important to trypanocidal activity; this fact was also considered crucial to exert antioxidant activity, because of their better insertion into lipophilic surfaces (Kato & Furlan, 2007). Several studies were reported about the antileishmaniasic activity of natural hydroquinone and related structures, however there are few results concerning their antichagasic potential. In this context, a prenylated dihydroquinone isolated from *Peperomia galioides* (Piperaceae) displayed a promising toxicity against *Leishmania brasiliensis*, *L. donovani* and *L. amazonensis*, but it was inactive on trypomastigote forms of *T. cruzi* (Mahiou et al., 1995; Mahiou et al., 1996).

The present investigation revealed that 3-geranylated hydroquinone and benzoic acid derivatives, isolated from leaves of *P. crassinervium*, showed potent trypanocidal activity. More extensive and detailed studies on the trypanocidal activity of natural and synthetic geranylated derivatives from Piperaceae species will be the target of future studies focusing on the understanding of the structural requirements of this group of natural products. Also, *in vitro* and *in vivo* unspecific toxicity should be investigated in order to evaluate their potential as new leader structures for the development of antichagasic drugs.

Acknowledgements

This work was funded by grants provided by the São Paulo State Research Foundation (FAPESP) (03/11524-9 and 05/57042-0) and was also supported by the BIOTA/FAPESP - Biodiversity Virtual Institute Program (www.biotasp.org.br). A.A. Lopes and S.N. López wish to thank FAPESP for the provision of scholarship and fellowship (03/01867-6 and 06/50086-5). M. Furlan, M.J. Kato and V.S. Bolzani are grateful to CNPq for research fellowships.

References

- Agrawal, P.K., & Bansal, M.C. (1989). Carbon-13 NMR of flavonoids. Amsterdam: Elsevier Science Publishers B.V.
- Baldoqui, D.C., Kato, M.J., Cavalheiro, A.J., Bolzani, V.S., Young, M.C.M., & Furlan, M. (1999). A chromene and prenylated benzoic acid from *Piper aduncum. Phytochemistry*, 51(7), 899–902.
- Camacho, M.D.R., Phillipson, J.D., Croft, S.L., Marley, D., Kirby, G.C., & Warhurst, D.C. (2002). Assessment of the antiprotozoal activity of *Galphimia glauca* and the isolation of new nor-secofriedelanes and nor-friedelanes. *Journal of Natural Products*, 65(10), 1457–1461.
- Danelutte, A.P., Lago, J.H.G., Young, M.C.M., & Kato, M.J. (2003). Antifungal flavanones and prenylated hydroquinones from *Piper crassinervium* Kunth. *Phytochemistry*, 64(2), 555–559.
- De Pasquale, R., Circosta, C., Occhiuto, F., De Rosa, S., & De Stefano, S. (1991). Pharmacological studies on terpenoids from marine sponges – analgesic and muscle-relaxant effects. *Phytotherapy Research: PTR*, 5(2), 49–53.
- Grael, C.F.F., Albuquerque, S., & Lopes, J.L.C. (2005). Chemical constituents of *Lychnophora pohlii* and trypanocidal activity of crude plant extracts and of isolated compounds. *Fitoterapia*, 76(1), 73–82.
- Jannin, J., & Villa, L. (2007). An overview of Chagas disease treatment. Memórias do Instituto Oswaldo Cruz, 102(1), 95–97.
- Jordão, C.O., Vichnewski, W., Souza, G.E.P., Albuquerque, S., & Lopes, J.L.C. (2004). Trypanocidal activity of chemical constituents of *Lychnophora salicifolia* Mart. *Phytotherapy Research: PTR, 18*(4), 332–334.
- Kato, M.J., & Furlan, M. (2007). Chemistry and evolution of the Piperaceae. *Chimie pure et appliquée*, 79(20), 529–538.
- Lago, J.H.G., Ramos, C.S., Casanova, D.C.C., Morandim, A.A., Bergamo, D.C.B., Cavalheiro, A.J., et al. (2004). Benzoic acid derivatives from *Piper* species and their fungitoxic activity against *Cladosporium cladosporioides* and *C. sphaerospermum. Journal of Natural Products*, 67(11), 1783–1788.
- Luize, P.S., Ueda-Nakamura, T., Dias Filho, B.P., Garcia Cortez, D.A., & Nakamura, C.V. (2006). Activity of Neolignans isolated from *Piper regnellii* (MIQ.) C. DC. var. *pallescens* (C. DC.) YUNCK against *Trypanosoma cruzi*. *Biological & Pharmaceutical Bulletin*, 29(10), 2126–2130.
- Mahiou, V., Roblot, F., Hocquemiller, R., Cave, A., Barrios, A., Fournet, A., et al. (1995). Piperogalin, a new prenylated diphenol from *Peperomia galioides*. *Journal of Natural Products*, 58(2), 324–328.
- Mahiou, V., Roblot, F., Hocquemiller, R., Cave, A., DeArias, A.R., Inchausti, A., et al. (1996). New prenylated quinones from *Peperomia galioides*. *Journal of Natural Products*, 59(7), 694–697.
- Navickiene, H.M.D., Alécio, A.C., Kato, M.J., Bolzani, V.S., Young, M.C.M., Cavalheiro, A.J., et al. (2000). Antifungal amides from *Piper hispidum* and *Piper tuberculatum*. *Phytochemistry*, 55(6), 621–626.
- Patrick, G.L. (1995). An Introduction to Medicinal Chemistry (1st ed.). Oxford: Oxford University Press.
- Pérez-Gutiérrez, E., Agrelo Salvatella, R., & Figueroa, R. (2006). Technical recommendation on Chagas' disease epidemiology and prevention, focussing its transmission as a disease transmitted by food (Revista da Sociedade Brasileira de Medicina Tropical, 39(5)) Brasilia: Sociedade Brasileira de Medicina Tropical.
- Rosas, L.V., Cordeiro, M.S.C., Campos, F.R.S., Nascimento, K.R., Januário, A.H., França, S.C., et al. (2007). *In vitro* evaluation of the cytotoxic and trypanocidal activities of *Ampelozizyphus amazonicus* (Rhamnaceae). *Brazilian Journal of Medical* and Biological Research, 40(5), 663–670.

- Silva, R.V.D., Navickiene, H.M.D., Kato, M.J., Bolzani, V.S., Méda, C.I., Young, M.C.M., et al. (2002). Antifungal amides from *Piper arboreum* and *Piper tuberculatum*. *Phytochemistry*, 59(5), 521–527.
- Wanke, S., Jaramillo, M.A., Borsch, T., Samain, M.S., Quandt, D., & Neinhuis, C. (2007). Evolution of Piperales—matK gene and trnK intron sequence data reveal lineage specific resolution contrast. Molecular Phylogenetics and Evolution, 42(2), 477–497.
- Yamaguchi, L.F., Lago, J.H.G., Tanizaki, T.M., Di Mascio, P., & Kato, M.J. (2006). Antioxidant activity of prenylated hydroquinone and benzoic acid derivatives from *Piper crassinervium* Kunth. *Phytochemistry*, 67(16), 1838–1843.